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Software Notice

PPLATE: a computer program for analysis of parallel-plate flow chamber experimental data

Yi Zhang, Sriram Neelamegham*

Bioengineering Laboratory, Department of Chemical Engineering, 906 Furnas Hall, State University of New York at Buffalo, Buffalo, NY 14260, USA

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13 1. Introduction

14 PPLATE is a computer program developed to analyze raw cell adhesion data obtained from the 1516parallel-plate flow chamber. It is based on a mathe-17 matical model described elsewhere (Zhang and Neelamegham, 2002a). The objective of this software is to 18 analyze raw data on cell rolling, firm-arrest and cell 1920transmigration to determine four frequency and prob-21ability parameters, namely, primary capture frequency, 22firm-arrest frequency, cell-cell capture probability and rolling-release frequency. These parameters are 23analogous to the "adhesion efficiency" used to quan-2425tify the rate of cellular aggregation in suspension. Overall, while the raw cell rolling and firm-arrest data 2627are a function of the physical features of the experiment system that control the rate of cell-substrate and 2829cell-cell collisions such as inlet cell concentration and wall shear stress, the frequency and probability 30 parameters are purely functions of the biological 31 32 adhesivity of cells and their response to fluid shear

forces. The program could find application in diverse33research areas that use the flow chamber device34including studies of leukocyte-endothelium interac-35tion, platelet adhesion at sites of vascular injury,36bacterial adhesion under fluid flow and cancer meta-37stasis.38

2. Description

The parallel-plate flow chamber is used to study 40 cell surface adhesion molecule function and receptor-41 ligand binding biophysics under physiologically rele-42vant hydrodynamic flow conditions (Gopalan et al., 431997; Goetz et al., 1999). Typical experiments quan-44 tify the interaction between the cells in suspension and 45the ligand-bearing flow chamber substrate in terms of: 46(i) rolling cell density, (ii) adhesion cell density and 47(iii) cell transmigration rate. Besides the biological 48properties of the cells, the physical features of the 49 system that control the number of cell-substrate and 50cell-cell collisions also influence these three meas-51ures. These physical features include, but are not 52limited to, the flow chamber size, wall shear stress, 53cell dimension and density, media density and inlet 54cell concentration. 55

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^{*} Corresponding author. Tel.: +1-716-645-2911x2220; fax: +1-716-645-3822.

E-mail address: neel@eng.buffalo.edu (S. Neelamegham).

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The program, PPLATE, is developed with the 56objective of separating the effects of the physical 5758features of the system listed above from the biological feature that control cellular adhesivity. It is 5960 based on a mathematical model described elsewhere 61 (Zhang and Neelamegham, 2002a,b). The objective of the model is to quantify four adhesion frequency 62and probability parameters for any given experimen-63 tal condition. These parameters are: (i) primary cap-64 ture frequency, (ii) cell-cell capture probability, (iii) 6566 firm-arrest frequency and (iv) rolling-release frequency. These parameters are purely measures of 67 the cellular adhesivity. While primary capture fre-68 quency quantifies the rate at which cells in the free 69 stream initiate rolling, the firm-arrest frequency is a 70measure of the rate of transition from rolling to firm-7172binding. The cell-cell capture probability quantifies the probability that cells in the free stream adhere 73onto the substrate following collision with previously 74recruited cells. Finally, the rolling-release frequency 7576quantifies the reversible release of cells from rolling 77 back into the free stream.

During each program run, the user is required to set 7879the inlet cell concentration and applied shear rate. 80 Following this, the user must decide to vary two of 81 the following variables/parameters over a defined range while keeping the remaining constant: dimen-82sionless position x^* , simulation time t, primary capture 83 frequency $\theta_{\rm fr}$, firm-arrest frequency $\theta_{\rm ra}$, cell-cell cap-84 ture probability θ_{cc} , rolling-release frequency θ_{rf} , and 85 transmigration time $t_{\rm TM}$. The reason why only two of 86 87 the parameters are varied is because this allows con-88 venient presentation of simulation results in the form of a two-dimensional matrix. The physical features of 89 the system, like the description of cell and media 90 91 attributes, and the flow chamber dimension are set to 92the default values listed in Table 1 of Zhang and Neelamegham (2002a). For a user with a flow chamber 93of different size or using cells with other dimensions, 94the default values can be readily varied by changing 95the parameters in the data file DATA.TXT, which 96 accompanies the software. This file can be saved in 97 98 text format using any software. The simulations per-99 formed here are not time-consuming, and it only takes ~ 10-15 s to simulate a 10-min flow chamber run on 100 a 366-MHz Pentium PC. We note that the run times 101102may vary depending on the nature of the simulation 103parameters chosen.

Simulation result was written into output file in text 104format. They can be read into many softwares includ-105ing Microsoft Excel and Notepad. The output file lists 106the default system parameters and adhesion frequen-107cies used in this run. This list of simulation parameters 108is followed by 3 two-dimensional data tables with the 109simulation results for: (i) rolling cell density, (ii) 110firmly adherent cell density and (iii) total transmi-111 grated cell density. Comparison of experimental data 112with simulation results allows estimation of the four 113frequency parameters for the given run as discussed 114elsewhere (Zhang and Neelamegham, 2002a). 115

3. Hardware/software requirements

The program is written in Digital Visual Fortran 117version 6.0 for IBM PC and compatibles. Recompi-118lation of the code requires linkage to the IMSL 119Numerical Library (Visual Numerics, San Ramon, 120CA). The executable version of the code (PPLA-121TE.EXE) has been tested on PCs running Windows 122operating system. Recompilation of the program on 123other operating systems (MacOS, UNIX) should also 124result in flawless execution of the code. 125

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4. Availability

The main program, PPLATE.EXE, is available 127from the journal's software archive (http://jim.mscs. 128mu.edu) under the directory pub/zhang via anony-129mous FTP. The FORTRAN source file PPLATE. 130FOR, input data file DATA.TXT, a word document 131containing sample program inputs (INPUT.DOC) and 132three sample output files that test the program under 133different conditions (RUN01.TXT, RUN02.TXT and 134RUN03.TXT) are also provided. We note that it is 135necessary that the file DATA.TXT be placed in the 136same directory as PPLATE.EXE during execution of 137the program. The output has been formatted for use/ 138manipulation by the speadsheet software Microsoft 139Excel. The output file is best opened by using the 140space-delimited feature of this software. The full manu-141 script describing the equations used in the software is 142published (Zhang and Neelamegham, 2002a). Copies 143of the program can also be obtained from: http:// 144 www.eng. buffalo.edu/~neel/pplate. 145

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146 References

- 147 Goetz, D.J., Greif, D.M., Shen, J., Luscinskas, F.W., 1999. Cell-
- 148cell adhesive interactions in an in vitro flow chamber. Methods149Mol. Biol. 96, 137–145.
- 150 Gopalan, P.K., Smith, C.W., Lu, H., Berg, E.L., McIntire, L.V.,
- 151 Simon, S.I., 1997. Neutrophil CD18-dependent arrest on inter-
- 152 cellular adhesion molecule 1 (ICAM-1) in shear flow can be
- activated through L-selectin. J. Immunol. 158, 367–375.
- Zhang, Y., Neelamegham, S., 2002a. An analysis tool to quantify
 the efficiency of cell tethering and firm-adhesion in the parallel
 plate flow chamber. J. Immunol. Methods (submitted for publication).
 156
- Zhang, Y., Neelamegham, S., 2002b. Estimating the efficiency of cell capture and arrest in flow chambers: study of neutrophil binding via E-selectin and ICAM-1. Biophys. J. 83 (in press).
 160